Corticostriatal Afferents Modulate Responsiveness to Psychostimulant Drugs and Drug-Associated Stimuli

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The medial prefrontal cortex (mPFC) and nucleus accumbens (NAc) are both integral components of the corticobasal ganglia–thalamic circuitry that regulates addiction-related behaviors. However, the role of afferent inputs from mPFC to NAc in these behaviors is unclear. To address this, we used a Cre-recombinase-dependent viral vector approach to express G_{i/o}-coupled DREADDs (designer receptors exclusively activated by designer drugs) selectively in mPFC neurons projecting to the NAc and examined the consequences of attenuating activity of these neurons on the induction of amphetamine sensitization and on drug taking and drug seeking during cocaine self-administration. Surprisingly, decreasing mPFC afferent activity to the NAc only transiently reduced locomotor sensitization and had no effect on drug taking during cocaine self-administration. However, inhibiting corticostriatal afferent activity during sensitization subsequently enhanced conditioned responding. In addition, this manipulation during drug self-administration resulted in slower rates of extinction and increased responding during drug prime-induced reinstatement—an effect that was normalized by inhibiting these corticostriatal afferents immediately before the drug prime. These results suggest that dampening cortical control over the NAc during drug exposure may lead to long-term changes in the ability of drugs and associated stimuli to drive behavior that has important implications for guiding treatments to prevent relapse.

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INTRODUCTION

Drug addiction is a debilitating neuropsychiatric disease characterized by a transition from controlled to uncontrollable drug taking and drug seeking and a high propensity to relapse, even after prolonged periods of drug abstinence. Chronic drug use produces adaptations within corticobasal ganglia–thalamic circuitry that are thought to underlie the behaviors that emerge during various stages of addiction (Kalivas and Volkow, 2011; Moussawi et al, 2011; Shiflett and Balleine, 2011). For example, morphological, electrophysiological, and neurochemical changes have been reported in both the striatum (ie, nucleus accumbens (NAc) and dorsal striatum), which is the central interface of this circuit, and the prefrontal cortex (PFC), which sends a strong glutamatergic input into the striatum (Berke and Hyman, 2000; Luscher and Malenka, 2011; Nestler, 2001; Russo et al, 2010; Schmidt and Pierce, 2010; Steketee and Kalivas, 2011). Although it has been postulated that a progressive reduction in PFC control over the striatum underlies many of the behaviors that contribute to addiction (Goldstein and Volkow, 2011; Kalivas et al, 2005; Feil et al, 2010; Steketee and Kalivas, 2011), until recently there has been little direct evidence for this, in part because the high degree of neuronal interconnectivity between the cortex and other regions of the corticobasal ganglia–thalamic circuit has made these studies difficult to conduct. In addition, the striatum receives innervation from multiple glutamatergic sources (thalamus, amygdala, and hippocampus) along with the cortex (McGeorge and Faull, 1989; Wall et al, 2013), and hence studies using traditional approaches, such as lesions or pharmacological blockade, have been unable to isolate which projections are critical for modulating striatal function in addiction-related behaviors. Thus, elucidating how PFC afferents to striatum in particular regulate behaviors associated with psychostimulant use is crucial for our understanding of the neural substrates that mark a transition to addiction as well as those that underlie relapse.

To investigate this, we used a novel Cre-recombinase (Cre-)–dependent, intersectional viral vector approach to express G_{i/o}-DREADDs (Designer Receptors Exclusively Activated by Designer Drugs) selectively in medial PFC (mPFC) afferents to NAc. DREADDs are only activated by clozapine-N-oxide (CNO); thus, this strategy allows for the transient and targeted activation of G_{i/o}-coupled signaling cascades in these neurons (Rogan and Roth, 2011). Activation of G_{i/o}-DREADDs by CNO decreases neuronal
activity primarily through a reduction in cAMP levels as well as activation of G protein-coupled inwardly rectifying potassium (GIRK) channels, resulting in membrane hyperpolarization and inhibition of neuronal firing (Armbruster et al., 2007; Ferguson et al., 2011, see Sternson and Roth, 2014 for review). Accordingly, these tools were used to examine how transiently decreasing activity of mPFC neurons that project to NAc through recruitment of inhibitory G protein-coupled signaling cascades affects the induction of psychomotor sensitization to amphetamine as well as the motivation to take drugs using a progressive ratio (PR) schedule of reinforcement in a cocaine self-administration paradigm and the motivation to seek drugs during drug prime-induced reinstatement. We hypothesized that decreasing activity of the cortical projections to NAc would block amphetamine sensitization as well as attenuate drug taking during cocaine self-administration and drug seeking during cocaine prime-induced reinstatement.

**MATERIALS AND METHODS**

**Experimental Strategy**

The overall experimental strategy was to use a combinatorial viral vector approach to express transgenes selectively in mPFC afferents to the NAc. This was achieved through injection of Cre-dependent adeno-associated virus (AAV) vectors into mPFC and a retrogradely transported canine adenovirus (CAV) expressing Cre into the NAc. Three separate experiments were then performed. In experiment 1, cocaine-induced c-Fos expression was measured in the mPFC and the NAc following activation of G<sub>iso</sub>-DREADDs in PFC to determine whether dampening PFC afferent input into the NAc was sufficient to decrease activity in NAc neurons. In separate experiments, the effect of dampening mPFC afferent input into the NAc was examined during the induction of amphetamine sensitization (experiment 2) and during cocaine self-administration (experiment 3) to determine whether corticostriatal projections modulate behaviors related to addiction. All experimental procedures were approved by the Seattle Children’s Research Institute institutional animal care and use committee and were conducted in accordance with National Institutes of Health (NIH) guidelines.

**Viral Vectors**

Cre-dependent AAV (serotype 5) vectors driven by the human synapsin promoter and expressing G<sub>iso</sub>DREADDs (AAV-hSyn-DIO-hM<sub>4</sub>Di-mCherry; hM<sub>4</sub>Di) or the control GFP (green fluorescent protein; AAV-hSyn-DIO-EGFP; GFP) were constructed by Dr Bryan Roth and obtained from the University of North Carolina viral vector core (titer of ~ 1 x 10<sup>9</sup> viral genomes/μl). CAV2-Cre (originally obtained from Dr Eric Kremer) was prepared in dog kidney (DK/E1-1) cells, purified by sucrose and CsCl gradient centrifugation steps, and resuspended in 1 x Hanks’ balanced saline solution at a titer of ~2.5 x 10<sup>10</sup> viral genomes/μl as described previously (Kremer et al., 2000).

**Experiment 1 (c-Fos)**

Male Long–Evans rats (n = 22, Charles River) weighing ~ 250–300 g were pair housed in a temperature- and humidity-controlled vivarium on a 12:12 h light–dark cycle and maintained on ad libitum food and water access. For viral-mediated gene transfer, rats were anesthetized with 2–4% isoflurane (Webster Veterinary Supply) and given meloxicam (0.2 mg/kg, s.c.) for pain management. Rats were monitored for at least 3 days following surgical procedures. Using standard stereotaxic procedures, 27-gauge stainless steel injectors were placed above the targeted brain regions. Coordinates from bregma (mm) for mPFC were A/P 3.2, M/L ±1.4, and D/V −3.5 from skull surface, and for NAc were A/P 1.7, M/L ±1.0, and DV −6.5. For assessment of cFos in the NAc, 2 μl of GFP was injected in one hemisphere of the mPFC, 2 μl of hM<sub>4</sub>Di was infused into the contralateral hemisphere of the mPFC, and 2 μl of CAV-Cre was infused bilaterally into the NAc over a 10-min period at a flow rate of 0.2 μl/min. Thus, each rat (n = 7) had GFP in one hemisphere and hM<sub>4</sub>Di in the other hemisphere, allowing for a within-subject design for this experiment. For assessment of cFos in the mPFC and in the basolateral amygdala (BLA), rats received bilateral injections of CAV-Cre into NAc and bilateral injections of GFP (n = 7) or hM4Di (n = 7) in the mPFC, allowing for a between-subject design. Accuracy of injection coordinates was confirmed by visualization of GFP or mCherry immunofluorescence in mPFC. At 20 days following viral infusions, rats were transported to a novel test environment and given an injection of CNO (3 mg/kg, i.p.; obtained from the NIH as part of the Rapid Access to Investigative Drug Program funded by the NINDS) followed 20 min later by an injection of cocaine (20 mg/kg, i.p.). After 2 h, rats were killed and brains were processed for immunohistochemistry; c-Fos cells were counted in the mPFC, the NAc, and the BLA. In addition, to assess whether the mPFC neurons projecting to the NAc also send collaterals to other brain regions within the corticostriatal circuit, GFP fluorescence was assessed in the NAc, the BLA, and the ventral hippocampus (VH).

**Experiment 2 (Sensitization)**

Male Sprague–Dawley rats (n = 52) underwent virulmediated gene transfer surgery as described for experiment 1, except that the surgical coordinates and injection volumes were modified. Coordinates from bregma (mm) for mPFC were A/P 2.8, M/L ±0.8, and D/V −4.5 from skull surface, and for NAc were A/P 1.8, M/L ±1.0, and DV −7.5, and rats received 1 μl of each virus at each site.

The psychomotor-activating effects of amphetamine were measured using locomotor activity boxes (San Diego Instruments). Briefly, at least 14 days following viral infusions, rats received four injections of amphetamine (2 mg/kg, i.p., Sigma) or vehicle (0.9% saline, i.p.) over a 7-day treatment period (one injection occurring every other day). At 20 min before each drug treatment, all rats received an injection of CNO (2 mg/kg, i.p.). Following injections of amphetamine or vehicle, rats were placed into the locomotor activity boxes where behavior was recorded for 90 min. After a 14-day withdrawal period, all rats underwent a challenge session. First, rats habituated to the locomotor chambers for
30 min. Then they received an injection of saline followed by
30 min of behavioral testing to assess for a conditioned
response. Finally, rats received a low-dose amphetamine
challenge (0.5 mg/kg, i.p.) in the absence of CNO pretreat-
ment followed by 90 min of behavioral testing to assess for
the persistence of sensitization. The number of cage
crossovers, defined as two consecutive beam breaks, was
used as an index of locomotor activity. Stereotypy ratings
were also assessed during testing using an adapted 9-point
rating scale (Dougherty and Ellinwood, 1983). Rats were
observed by an experimenter blind to the experimental
conditions for 30 s every 5 min during the test sessions
and were given a stereotypy rating during each observation
(1, asleep; 2, inactive; 3, normal in-place activity; 4, normal,
alert, active; 5, hyperactive; 6, slow patterned stereotyped
behaviors; 7, fast patterned stereotyped behaviors; 8,
restricted stereotyped behaviors; 9, dyskinetic reactive).
There were four groups used in the psychomotor sensitiza-
tion experiments: hM4Di rats injected with amphetamine
(n = 13) or saline (n = 12) and GFP rats injected with
amphetamine (n = 13) or saline (n = 12). Two rats were
excluded from the analysis because virus expression was
outside of mPFC.

**Experiment 3 (Self-Administration)**

Male Long–Evans rats (n = 35) underwent viral-mediated
gene transfer surgery as described in experiment 1, with rats
receiving bilateral injections of either GFP (n = 7) or hM4Di
(n = 15) into the mPFC and bilateral injections of CAV-Cre
into the NAc. Following recovery, indwelling jugular
catheters were implanted as previously described
(Kerstetter et al., 2008). Briefly, catheters were inserted into
the right jugular vein and connected to a back-mounted port.
Catheters were flushed daily with Timentin antibiotic
(20 mg/kg, i.v.; Butler Schein) and catheter patency was
verified periodically with methohexital sodium infusions
(10 mg/ml i.v.; Eli Lilly).

At least 14 days following viral infusions, rats were trained
to self-administer cocaine (0.75 mg/kg/infusion in 100 μl of
0.9% sterile saline administered over 4 s; obtained from
the National Institute on Drug Abuse) during their light cycle on
a fixed interval 20 s (FI:20) schedule of reinforcement. Each
cocaine infusion was paired with a 5 s light and tone
stimulus. Self-administration sessions lasted 2 h and took
place 5 days per week in sound-attenuated operant
conditioning chambers (Med Associates).

After rats had met self-administration criteria (minimum of
6 training sessions with 10 or more cocaine infusions
earned for 3 consecutive sessions), they began PR sessions
during which the response requirement to earn a cocaine
infusion increased after each infusion earned. The response
requirement progression followed that of Richardson and
Roberts (1996) and was as follows: 1, 2, 4, 6, 9, 12, 15, 20, 25,
32, 40, 50, 62, 77, 95, 118, 145, 178, 219, 268, 328, 402, 492,
603. PR sessions were limited to 2 h because of the duration
of CNO treatment (which was given during the testing
phase). Once behavior had stabilized (≤2-step change in the
last response requirement completed in a session over 2
consecutive sessions), rats were administered vehicle
(6% DMSO in sterile water, i.p.) 20 min before PR testing
for three sessions to obtain a baseline response. Rats were
then administered CNO (3 mg/kg, i.p.) 20 min before PR
testing for three sessions. Following testing, rats underwent
three additional PR sessions in the absence of any
pretreatment to determine whether CNO treatment had
any lasting effects on PR responding.

A subset of rats (GFP: n = 7; hM4Di: n = 5) then underwent
extinction of operant responding until criterion was met
(a minimum of 7 sessions with at least 2 consecutive sessions
of ≤25 active lever responses). During the 2 h extinction
sessions, levers were extended but responding did not have
any programmed consequences (i.e., no infusions or
light/tone cue were given). Rats then received vehicle
treatment immediately before being placed into the operant
chamber to establish a baseline of responding. Rats then
underwent two cocaine prime-induced reinstatement tests
(10 mg/kg, i.p.) with either DMSO or CNO (3 mg/kg, i.p.)
given 20 min before the cocaine injections. Cocaine
injections were given immediately before reinstatement testing,
and levers were extended for these sessions but responding
did not have any programmed consequences. These
pretreatments were given in a counterbalanced manner and
rats received additional extinction training sessions following
the first reinstatement test (see Figure 4 for illustration
of experimental design). In all, 13 rats were excluded from the
experiments because 4 rats had injection sites outside of the
mPFC, 7 rats failed to acquire cocaine self-administration,
and 2 rats were outliers from the behavioral data set (> 2 SD
away from the mean).

**Immunohistochemistry**

Rats were anesthetized with Beuthanasia-D (Schering-
Plough) and perfused transcardially with 1 × PBS (pH 7.4),
followed by 4% paraformaldehyde (PFA). Brains were
extracted, post-fixed in 4% PFA overnight, and stored in
1 × PBS. Floating sections (40–60 μm) were washed in 0.5%
Triton-X/PBS for 10 min, blocked in 5% normal goat serum
or 7.5% normal donkey serum (NS)-0.25%Triton-X/PBS for
2 h, and incubated in 2.5–5% NS-0.25% Triton-X/PBS con-
taining antibodies to GFP (1 : 400, Millipore), mCherry
(1 : 400, Clontech), or c-Fos (1 : 400, Santa Cruz) with gentle
agitation for 72 h. Next, sections were rinsed 4 times in
PBS and incubated in species-appropriate Alexa 488 (green),
Alexa 568 (red), or Alexa 647 (far red)-conjugated secondary
antibodies (1 : 400, Invitrogen) for 2 h. Sections were washed
2 times in PBS, mounted on slides, and coverslipped with
Vectashield mounting medium with DAPI (Vectorlabs).
Z-stack images were captured with a Zeiss confocal
microscope and compressed into a single plane before
quantification. c-Fos+ cells in the mPFC, NAc, and BLA
were counted and averaged across 3–4 sections for each rat
using ImageJ software (NIH).

**Data Analysis**

All analyses consisted of planned (a priori) comparisons. Group
differences in crossovers, stereotypy ratings, active
and inactive lever presses, and number of earned infusions
were tested using two-way analysis of variance (ANOVA)
with repeated measures when applicable, followed by
Bonferroni’s post hoc tests. Differences in the number of
c-Fos+ cells were tested using a paired t-test for the NAc and
unpaired t-tests for the mPFC and the BLA. For all comparisons, \( \alpha \leq 0.05 \). Data are graphed as mean ± SEM.

RESULTS

The Intersectional Viral Vector Approach Produces Transgene Expression Primarily in mPFC Neurons Projecting to the NAc

A Cre-dependent intersectional vector approach was used to express hM4Di receptors in mPFC neurons projecting to the NAc in order to selectively and transiently decrease activity of these afferents (Figure 1a and b). Viral expression in the mPFC was largely confined to the cingulate and prelimbic regions (Figure 1). Although transgene expression was induced by retrograde infection of Cre from NAc neurons, it is possible that the DREADD-expressing mPFC neurons send collaterals to other regions. In order to confirm specificity of the intersectional approach, we examined fluorescence expression in mPFC terminals in the NAc, as well as the BLA and the VH, the two output regions of the mPFC that also send glutamatergic inputs into the NAc. We observed a strong amount of fluorescence in the NAc (Figure 1c), with some fluorescence in the BLA (Figure 1d) and no fluorescence in the VH (Figure 1e), suggesting that DREADD receptor expression was occurring primarily in mPFC neurons that were projecting selectively to the NAc.

Decreasing Activity of mPFC Afferents to NAc Reduces Cocaine-Induced c-Fos in Both PFC and NAc Neurons

It is well established that activation of hM4Di receptors by CNO decreases neuronal activity primarily through a reduction in cAMP levels as well as activation of GIRK channels (Armbruster et al., 2007; Ferguson et al., 2011, see Sterenson and Roth, 2014 for review). This effect has been observed in glutamatergic pyramidal neurons of the cortex, the population of neurons that expressed hM4Di receptors in the present set of experiments (Katzel et al., 2014; Kozorovitskiy et al., 2012; Robinson et al., 2014). Using cocaine-induced stimulation of the immediate early gene c-Fos as a marker of neuronal activity, we found that consistent with these studies, activation of hM4Di receptors in mPFC significantly decreased the number of cocaine-evoked c-Fos+ cells in mPFC by ~30% compared with the GFP controls (Figure 2a and b; \( t_{6} = 2.68, P = 0.02 \)). In order to extend these findings, we examined whether the manipulation in mPFC was sufficient to alter activity of the striatal neurons receiving mPFC input. Indeed, we found that activation of hM4Di receptors in mPFC significantly decreased the number of cocaine-evoked c-Fos+ cells in NAc by ~32% compared with the GFP control hemisphere (Figure 2c and d; \( t_{6} = 0.56, P = 0.01 \)), suggesting that decreasing activity of mPFC projections to NAC reduces the neuronal activity of its downstream targets. In contrast, CNO-induced activation of hM4Di receptors in mPFC had no effect on cocaine-evoked c-Fos+ cells in the BLA (Figure 2e and f; \( t_{13} = 0.9, P = 0.38 \)), suggesting that the manipulation was selective for altering activity in mPFC to NAc neurons.

Decreasing Activity of mPFC Afferents to NAc Alters the Induction of Amphetamine Sensitization

We used the intersectional DREADD viral vector approach to examine whether decreasing activity of mPFC afferents to NAc during amphetamine administration would be sufficient to block the induction of this progressive and persistent form of drug-induced behavioral plasticity. We found that CNO-induced activation of hM4Di receptors had no effect on locomotor activity following saline injections (Figure 3a; main effect of virus \( F_{5, 22} = 4.97, P = 0.04 \); main effect of session \( F_{2, 66} = 6.12, P = 0.001 \); no interaction between session and virus \( F_{5, 66} = 1.35, P = 0.27 \)), suggesting a lack of effect of this manipulation on baseline activity. Although both locomotor and stereotyped responses to amphetamine increased over sessions in GFP and hM4Di rats
Aperf表情达意的药物。最后，所有老鼠都接受了注射程序，但没有最后开发条件反射的注射的程序。有趣的是，hM4Di 老鼠在以前接受过安非他命的注射，显示增强的活动在mPFC 脑区。根据CNO 以评估减少mPFC 向NAc 的投射的活动。To determine whether decreasing activity of mPFC afferents to NAc alters drug taking in a cocaine self-administration paradigm, rats were trained to self-administer cocaine first on a FI20 schedule and then on a PR schedule of reinforcement until stable baselines were achieved (Figure 4). During the last three PR training sessions before testing, both GFP and hM4Di rats made significantly more active lever responses compared with inactive lever responses (Figure 4a; main effect of lever, hM4Di: F1,28 = 51.70, P < 0.0001; GFP: F1,12 = 50.73, P < 0.0001) but there were no differences in lever responding across sessions (Figure 4a; no main effect of session, hM4Di: F2,56 = 1.57, P = 0.38; GFP: F2,24 = 0.37, P = 0.70; no interaction between lever and session, hM4Di: F2,56 = 1.00, P = 0.38; GFP: F2,24 = 0.47, P = 0.63). In addition, there were no differences in either active lever responses (Figure 4a; no main effect of virus F1,20 = 0.20, P = 0.66; no main effect of session F2,40 = 0.03, P = 0.97; and no interaction between virus and session F2,40 = 1.42, P = 0.25) or number of earned infusions completed (Figure 4b; no main effect of virus F1,20 = 0.14, P = 0.71; no main effect of session F2,40 = 0.30, P = 0.74; and no interaction between virus and session F2,40 = 0.56, P = 0.59) between GFP and hM4Di rats, suggesting that both groups had acquired the same stable levels of cocaine self-administration. Both groups then received vehicle pretreatment for three sessions, followed by CNO pretreatment for three sessions and three additional baseline sessions (data averaged across sessions). There were no differences between groups in active lever responses (Figure 4a; no main effect of virus F1,20 = 1.03, P = 0.32; no main effect of drug pretreatment F2,40 = 0.53, P = 0.60; and no interaction between virus and drug pretreatment F2,40 = 1.90, P = 0.16) or number of earned infusions (Figure 4b; no main effect of
Motivation for Seeking Drugs

Decreasing Activity of mPFC Afferents to NAc Alters the Development of Amphetamine Sensitization.

We used a Cre-dependent intersectional viral vector approach to express Gi/o-DREADDs in mPFC neurons that project to the NAc. Consistent with the known mechanism of hM4Di receptor activation, which is to reduce excitability of the cells expressing the Gi/o-DREADDs (Armbruster et al., 2007; Ferguson et al., 2011, see Sterpenich and Roth, 2014 for review), we found that CNO-induced receptor activation reduced cocaine-evoked c-Fos in the region of viral expression. In order to assess the specificity of this approach, we compared immunofluorescence in GFP-expressing rats in three terminal regions of the mPFC—the NAc, the BLA and the VH. Although the densest amount of fluorescence was observed in the NAc, fluorescence was also apparent in the BLA and the VH. The densest amount of fluorescence was observed in the NAc, suggesting that the corticostriatal neurons that were expressing Gi/o-DREADDs may send axon collaterals to other regions. However, it is also possible that the signal in the BLA was simply because of axons passing through the region. In line with this idea, we found that activation of hM4Di receptors in mPFC attenuated cocaine-evoked c-Fos in NAc neurons but not in BLA neurons. Cocaine-induced activation of c-Fos in striatal cells is modulated by glutamate (Harlan and Garcia, 1998); thus, it is likely that the reduction in c-Fos that we observed following hM4Di receptor activation of corticostriatal neurons was because of a decrease in glutamatergic stimulation of the NAc neurons. In addition, these results indicate that the intersectional

during the induction phase. Unexpectedly, hM4Di rats that had received CNO treatment during PR testing made significantly more active lever responses during the first 2 days of extinction training compared with the GFP controls (Figure 4c; main effect of virus $F_{1,10} = 12.90, P = 0.005$; main effect of session $F_{6,60} = 50.07, P < 0.0001$; and interaction between virus and session $F_{6,60} = 5.76, P < 0.0001$), suggesting an impairment in their extinction learning. In addition, although both groups reached extinction criteria by 10 sessions, hM4Di rats made significantly more active lever responses following a priming injection of cocaine compared with the GFP controls (Figure 4d; main effect of virus $F_{1,10} = 2.67, P = 0.03$; main effect of drug treatment $F_{2,20} = 38.96, P < 0.0001$; and interaction between virus and drug treatment $F_{2,20} = 9.19, P = 0.002$), indicating a greater level of reinstatement. Interestingly, CNO treatment immediately before the cocaine prime significantly attenuated reinstatement in the hM4Di rats (Figure 4d; main effect of virus $F_{1,10} = 9.56, P = 0.01$; main effect of pretreatment $F_{2,20} = 38.96, P < 0.0001$; and interaction between virus and pretreatment $F_{2,20} = 9.19, P = 0.002$). These effects were not due to indiscriminate alterations in activity, as inactive lever responses did not differ between groups during extinction training (Figure 4c; no main effect of virus $F_{1,10} = 3.70, P = 0.08$; no interaction between virus and session $F_{6,60} = 0.38, P = 0.89$) or during cocaine prime-induced reinstatement (no main effect of virus $F_{1,10} = 0.29, P = 0.60$; no main effect of drug pretreatment $F_{1,10} = 3.94, P = 0.08$; and no interaction between virus and drug pretreatment $F_{1,10} = 1.26, P = 0.29$), and CNO had no effect on cocaine prime-induced reinstatement in the GFP controls (Figure 4d; $P = 0.80$).

Figure 3

Amphetamine sensitization. Decreasing activity of mPFC afferents to NAc alters the development of amphetamine sensitization. CNO-mediated activation of hM4Di in corticostriatal afferents significantly decreased locomotor sensitization to amphetamine as measured by crossovers (a, $*P < 0.05$ versus GFP group), but had no effect on stereotypy ratings (b). Decreasing activity of mPFC afferents to NAc during the induction phase of sensitization significantly increased conditioned responding as measured by crossovers during a saline challenge (c, $*P < 0.05$ versus hM4Di group given saline during the induction phase), but had no effect on the persistence of locomotor sensitization as measured by crossovers during a low-dose amphetamine challenge (0.5 mg/kg, i.p.) in the absence of CNO pretreatment (d, $*P < 0.05$ versus respective control groups given saline during the induction phase). Data represent mean ± SEM. Black symbols/shading (hM4Di) and white symbols/shading (GFP) represent groups that received amphetamine treatment during the induction phase. Dark gray symbols/shading (hM4Di) and light gray symbols/shading (GFP) represent groups that received saline treatment during the induction phase. S: saline during the induction phase, A: amphetamine during the induction phase. N = 11–17/group.

**DISCUSSION**

We used a Cre-dependent intersectional viral vector approach to express Gi/o-DREADDs in mPFC neurons that project to the NAc. Consistent with the known mechanism of hM4Di receptor activation, which is to reduce excitability of the cells expressing the Gi/o-DREADDs (Armbruster et al., 2007; Ferguson et al., 2011, see Sterpenich and Roth, 2014 for review), we found that CNO-induced receptor activation reduced cocaine-evoked c-Fos in the region of viral expression. In order to assess the specificity of this approach, we compared immunofluorescence in GFP-expressing rats in three terminal regions of the mPFC—the NAc, the BLA and the VH. Although the densest amount of fluorescence was observed in the NAc, c-Fos fluorescence was also apparent in the BLA, suggesting that the corticostriatal neurons that were expressing Gi/o-DREADDs may send axon collaterals to other regions. However, it is also possible that the signal in the BLA was simply because of axons passing through the region. In line with this idea, we found that activation of hM4Di receptors in mPFC attenuated cocaine-evoked c-Fos in NAc neurons but not in BLA neurons. Cocaine-induced activation of c-Fos in striatal cells is modulated by glutamate (Hanson and Garcia, 1998); thus, it is likely that the reduction in c-Fos that we observed following hM4Di receptor activation of corticostriatal neurons was because of a decrease in glutamatergic stimulation of the NAc neurons. In addition, these results indicate that the intersectional
approach was selective for modulating corticostriatal afferent activity.

Next, we examined the consequences of transiently decreasing activity in corticostriatal neurons during amphetamine administration on psychomotor sensitization. Given that previous work has demonstrated a role for the mPFC in regulating sensitization (Wolf, 1998; Vandershuren and Kalivas, 2000; Tzschentke and Schmidt, 2003; Kalivas, 2004; Steketee and Kalivas, 2011), and many of the sensitization-related neurobiological changes that occur in the NAc are glutamate dependent (Kalivas et al., 2005; Steketee and Kalivas, 2011), we hypothesized that selectively dampening mPFC activity to the NAc would decrease the induction of sensitization. Consistent with this hypothesis, decreasing activity of mPFC afferents to NAc during repeated amphetamine administration attenuated locomotor sensitization without altering stereotyped responses that are thought to be regulated by the dorsal striatum (Staton and Solomon, 1984). Unexpectedly, locomotor sensitization appeared equivalent between groups during the challenge phase. There are several possible explanations that could account for these results. First, the observed effects during the induction phase could be due to a decrease in the expression of sensitization; this is unlikely to be the case as differences in the level of sensitization were only observed during the last treatment session. Second, it is possible that our manipulation initially decreased the induction of sensitization, but the underlying neurobiological changes that occur in corticostriatal circuits during withdrawal permitted sensitization to develop normally over the long term. Third, it may be that decreasing mPFC afferent activity to the NAc did, in fact, permanently disrupt the induction of sensitization, and the response during the amphetamine challenge was a reflection of other processes that were altered.

Figure 4  Cocaine self-administration. Decreasing activity in mPFC afferents to NAc modulates drug seeking. There were no differences between groups in active (circles) and inactive (squares) lever responses (a, left panel) and number of earned infusions (b, left panel) during baseline progressive ratio (PR) cocaine self-administration (last 3 sessions before testing). CNO-mediated activation of hM₄Di in corticostriatal afferents had no effect on lever responses (a, right panel) or number of earned infusions (b, right panel). Rats that had hM₄Di activation during PR showed a significant increase in the number of active lever responses during the first 2 sessions of extinction training (c, *P<0.05 versus GFP group) as well as following a cocaine prime (d, *P<0.05 versus GFP group). CNO pretreatment before a cocaine prime significantly attenuated active lever responding only in the hM₄Di group (d, #P<0.05 versus V-treated hM₄Di group). Data represent mean ± SEM. Black symbols in line graphs represent the hM₄Di group and white symbols represent the GFP controls. *Note that pretreatments (V or C) were counterbalanced across rats during reinstatement tests. SAL: saline injection; COC: cocaine injection; V: vehicle pretreatment; C: CNO pretreatment. N=5–7 group.
during the DREADD manipulation, such as an increase in conditioned responding. In support of this last idea, we found that attenuating activity in cortical projections to NAc during the induction of a conditioned response to the injection procedure as measured during the saline challenge—an effect not seen in the controls.

Nonetheless, these results suggest that direct modulation of NAc activity by the mPFC is unlikely to be a critical node in the modulation of behavioral sensitization. Instead, it is likely that the mPFC exerts its effects on NAc indirectly through regulation of other NAc inputs, such as VTA, to modulate sensitization, and other sources of glutamate (ie, amygdala, hippocampus, or thalamus) must be responsible for direct modulation of the NAc. Consistent with this idea, it was recently demonstrated that optical inhibition of VH inputs into the NAc shell was sufficient to reduce the development of locomotor sensitization to cocaine (Britt et al, 2012). In addition, decreasing activity of the BLA via G\textsubscript{i/o}-DREADDs not only attenuated the development of cocaine sensitization but also blocked cocaine-induced increases in the frequency of miniature excitatory postsynaptic currents in NAc neurons, suggesting that NAc activity is directly modulated by amygdala afferents (MacAskill et al, 2014). However, it should be noted that in the present set of experiments DREADD receptors were primarily expressed in the cingulate and prelimbic regions of the mPFC; thus, we cannot rule out that targeting a larger region of the PFC and/or more medial aspects could lead to a different behavioral outcome. This is unlikely to be the case, however, as previous work has demonstrated that lesions of or pharmacological manipulations to this region of mPFC are sufficient to modulate sensitization (Tzschentke and Schmidt, 2003).

Finally, we examined the consequences of transiently decreasing activity in corticostriatal neurons on drug taking during cocaine self-administration and on drug prime-induced reinstatement following extinction. Although this manipulation had no effect on on-going drug use during self-administration, inhibiting mPFC afferent activity to the NAc immediately before drug prime decreased reinstatement. This finding is consistent with recent studies that found that optogenetic inhibition of prelimbic PFC fibers in the NAc core also blocks both cocaine prime-induced reinstatement and cocaine-plus-cue-induced reinstatement (Stefanik et al, 2013, 2015). In addition, it is consistent with recent work demonstrating that reversing silent synapse remodeling (that occurred during withdrawal from cocaine self-administration) in the NAc core by optical stimulation of PFC inputs inhibited cue-induced cocaine seeking (Ma et al, 2014). Finally, these results are in line with the idea that heightened PFC activation underlies the enhanced responsiveness seen to drugs and drug-related cues during relapse (Feil et al, 2010; Goldstein and Volkow, 2011).

Although decreasing cortical activity to NAc neurons did not alter drug-taking behavior during cocaine self-administration, it did have a large impact on subsequent responsiveness to the environmental stimuli associated with drug administration and to the drug itself. Specifically, decreasing activity in these neurons during cocaine self-administration led to a slower rate of extinction and an increase in active lever pressing during drug prime-induced reinstatement compared with controls. Together with the enhanced conditioned response seen following amphetamine sensitization, these results suggest that cortical inputs into NAc may be modulating the strength of associations between drugs and the circumstances surrounding drug administration (ie, ‘set and setting’) and are consistent with imaging studies in human addicts as well as in preclinical animal models that have found that, relative to controls, extensive psychostimulant use results in a hypoactive PFC at baseline but heightened PFC activation to both the drug and drug-related cues (Goldstein and Volkow, 2011).

Nonetheless, it is perhaps counterintuitive that dampening mPFC afferent activity to NAc during initial drug use would subsequently lead to the development of a conditioned response, as well as the delayed extinction and enhanced responding during drug prime-induced reinstatement (all of which occurred in the absence of DREADD receptor activation). However, although much work has focused on the brain regions and circuits that regulate how drug-associated stimuli drive reinstatement of cocaine-seeking behavior (Everitt and Robbins, 2005; LaLumiere et al, 2012; Marchant et al, 2014), surprisingly little is known about the neural mechanisms that underlie how drugs initially change the incentive value of the stimuli that become associated with drug use. Our results indicate that increasing G\textsubscript{i/o} signaling in corticostriatal neurons could help drive the strength of these associative processes. In addition to reducing cAMP activity and activating GIRK channels (Sternson and Roth, 2014), induction of G\textsubscript{i/o} cascades produces a slow and sustained increase in ERK/MAPK (extracellular signal-regulated kinase) signaling through a β-arrestin-mediated pathway (Reiter and Lefkowitz, 2006; Wettscueureck and Offermanns, 2005). ERK/MAPK signaling cascades are important modulators of long-term alterations in neuronal plasticity and memory formation (Giovannini, 2006; Sweatt, 2004), and it is noteworthy that the changes in responsiveness to the stimuli associated with drug use took a period of time to develop, as they were not evident during the induction of sensitization (that appeared blunted) or in the three baseline self-administration sessions that followed CNO treatment. Thus, β-arrestin-mediated recruitment of ERK/MAPK cascades during drug use is one possible mechanism that could facilitate the strengthening of associations between the drug and the stimuli. Nonetheless, the striatum receives innervation from multiple glutamatergic sources (thalamus, amygdala, and hippocampus) along with the cortex (McGeorge and Faull, 1989; Wall et al, 2013), and it is not yet known whether these inputs work in concert or in opposition to regulate NAc neurons and subsequent behavioral output. Thus, it is also possible that dampening mPFC activity allowed for the information that is carried from these other inputs to have a larger impact on NAc neuron function, thereby facilitating the development of associations between the drug and the ‘set and setting’ surrounding drug use.

In summary, this work helps to elucidate how mPFC afferents to NAc, in particular, regulate addiction-related behaviors and govern the processes that contribute to relapse. Our results suggest that rather than modulating the maintenance of ongoing behaviors as previously thought, these afferent connections from mPFC to NAc may instead be key for shaping the associations between drugs and the
stimuli surrounding drug use, as well as in reinstatement of drug-seeking behaviors. Given that relapse to drugs following exposure to drug-associated stimuli is one of the most insidious facets of addiction, particularly because it is such a persistent phenomenon (Frawley and Smith, 1992; Jones et al, 2003), this work has important clinical implications as it suggests that the mPFC to NAc input would be a promising target for therapeutic intervention.

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AUTHOR CONTRIBUTIONS
KAK, AMW, KGN, and ED performed the behavioral and immunohistochemical experiments. JPN provided the CAV-Cre. KAK, AMW, and SMF designed the overall study and wrote the manuscript. All authors contributed to data interpretation and manuscript editing.

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